A NEW FLUORIDOLYSABLE ANCHORING LINKAGE FOR ORTHOGONAL SOLID-PHASE PEPTIDE SYNTHESIS: PREPARATION AND PROPERTIES OF THE <u>N</u>-(3 OR 4)-[[[(4-HYDROXYMETHYL)-PHENOXY-<u>t</u>-BUTYLPHENYL]SILYL]PHENYL]PENTANEDIOIC ACID, MONOAMIDE (PBS) HANDLE¹⁻³ Daniel G. Mullen and George Barany^{*}

University of Minnesota, Minneapolis, Minnesota 55455

The synthesis and characterization of a new silicon-containing handle for use in solid-phase peptide synthesis is described. The anchoring linkage derived from this new handle, when treated with fluoride (1.0 equiv.) for 5 min at 25 °C, releases peptides as their free acids in essentially quantitative yields and without racemization.

As part of a program⁴ to develop mild chemical methods for solid-phase peptide synthesis⁵, we require new orthogonally⁶ cleavable handles^{7,8} for attaching chains to the support. Silyl ether-containing linkages were considered as attractive candidates because of the possibility for facile cleavage under neutral conditions of fluoridolysis^{9,10}. We report here the preparation of the new protected <u>N</u>-(3 or 4)-[[4-(aminoacyloxymethyl)phenoxy-t-butylphenyl]silyl]phenyl-N'-resinylpentanedioic acid, diamide (1) which is designed so



that rapid silicon-oxygen⁹ bond fission with one equivalent of fluoride at 25 °C is followed directly by a 1,6-elimination mechanism¹¹ leading to a free peptide acid (see above diagram). We further show how this anchoring linkage, which because of the <u>p</u>-alkoxybenzyl ester¹² moiety can also be cleaved by trifluoroacetic acid at 25 °C, is compatibly applied with \underline{N}^{α} -9-fluorenylmethyloxycarbonyl (Fmoc)¹³ and \underline{N}^{α} -dithiasuccinoyl (Dts)⁴ amino acids for stepwise solid-phase synthesis of a simple model peptide.

Resin derivative 1 was synthesized in twelve steps (see Scheme)¹⁴ starting from commercially available <u>t</u>-butylchlorodiphenylsilane (2). Chlorosilane 2 could be cleanly hydrolysed [KOH (1.1 equiv.) in H_20 —MeOH—Et₂O (4:1:6), 25 °C, 12 h] to the relatively inert corresponding silanol, which was carefully nitrated (-23 °C, 5 h) with ammonium nitrate (1.15 equiv.) according to Crivello¹⁵. The desired mononitrosilanol 3 was isolated

apart from unreacted starting material and the dinitro compound by silica gel chromatography $[Et_20-hexane (3:17)]$. It was established by two dimensional COSY NMR at 300 MHz that the <u>meta</u> and <u>para</u> substituted isomers both formed (4:1 ratio). This ratio of isomers carried over for subsequent products. Next, a new method was developed using oxalyl chloride (4 equiv.) and DMAP (1.5 equiv.) in CH_2Cl_2 (25 °C, 36 h) to transform the silanol back to a chlorosilane. Subsequent reaction (12 h, under N₂) with <u>p</u>-hydroxybenzaldehyde (1.1 equiv.) in the presence of <u>N</u>,<u>N</u>-diisopropylethylamine (1.5 equiv.) in dry CH_2Cl_2 gave silyl ether 4.

The aldehyde of **4** was then reduced (under N₂) using borane-THF complex (1.0 equiv.) in ethyl ether (25 °C, 1 h) to provide a benzylic alcohol function which was protected¹⁶ as the tetrahydropyranyl ether. These steps gave **5**, which required modification before attachment to polymeric supports was possible. Consequently, the nitro group of **5** was reduced¹⁷ by catalytic transfer hydrogenation [NH₄O₂CH (9.0 equiv.), Pd/C] in methanol (25 °C, 2 h) to give an amine, which was then acylated with glutaric anhydride¹⁸ (1.1 equiv.) in chloroform (25 °C, 12 h). The product of these steps was isolated as the solid dicyclohexylammonium salt **6**, mp 101 °C, and could be converted back to the free acid by neutralization with Dowex-50 ion-exchange resin. The acid was then esterified with 2,4,5-trichlorophenol (1.1 equiv.) using as the coupling agent N.M'-dicyclohexylcarbodiimide in CH₂Cl₂ (25 °C, 12 h), and finally deprotection with pyridinium tosylate¹⁶ (0.1 equiv.) in ethanol (50 °C, 4 h) gave the new silicon-functionalized handle **7**. The "preformed handle" approach^{4b,8a,d,e}

for attachment of amino acids to the polymeric support was then applied; thus, suitably protected amino acids were esterified to 7 using $\underline{N},\underline{N}$ -dimethylformamide dineopentyl acetal^{8d} as the condensing agent. The amino acyl handle (3.0 equiv.) was then quantitatively attached onto an aminomethylcopoly(styrene-1%-divinylbenzene) resin (0.19 meq/gm) by overnight reaction (25 °C) in the presence of 1-hydroxybenzotriazole (3.0 equiv.) in $\underline{N},\underline{N}$ -dimethyl-formamide.

Resin 1 was the starting point for several syntheses of the model tetrapeptide \underline{L} -leucyl- \underline{L} -alanylglycyl- \underline{L} -valine. Both Fmoc and Dts-amino acids were incorporated under standard conditions of this laboratory^{4b,8d} for DCC-mediated coupling. Upon completion of chain assembly, the peptide was rapidly cleaved by tetrabutylammonium fluoride (1.0 equiv.) in $\underline{N}, \underline{N}$ -dimethylformamide at 25 °C in 5 min. The cleavage yields were $\geq 97\%$ based on amino acid remaining on the resin, and the purities were $\geq 98\%$ as judged by ion-exchange chromatography. In one synthesis, the last amino acid was introduced with \underline{t} -butyloxycarbonyl (Boc) for \underline{N}^{α} -amino protection, and the orthogonal fluoridolysis conditions released the protected peptide acid Boc-Leu-Ala-Gly-Val-OH^{19,20}. Lastly, the issue of racemization was addressed by applying the Manning-Moore assay²¹ on the dipeptide alanylvaline sequence assembled on the resin and cleaved by fluoridolysis. Only the $\underline{L}, \underline{L}$ -diastereomer was observed (sensitivity limit 0.05%).

References and Notes

- 1. A preliminary report on this topic was presented by D.G.M. at the 187th National American Chemical Society Meeting in St. Louis, MO, April 8-13, 1984. Recently, we learned of independent work carried out by R. Ramage and colleagues at the University of Edinburgh, Scotland and reported at the Nineteenth European Peptide Symposium, Porto Carras-Chalkidiki, Greece, August 31-September 5, 1986. These workers prepared a resinbound variant of the Tmse group (see ref. 10a) and used it to prepare a model peptide which was cleaved by fluoridolysis.
- Financial support from the National Institutes of Health (GM 28934 and AM 01099) is gratefully acknowledged. D.G.M. thanks the Henkel Corporation for a graduate fellowship, 1984-1985.
- 3. Abbreviations used are: Boc, t-butyloxycarbonyl; DCC, N,N'-dicyclohexylcarbodiimide; DCHA, N,N-dicyclohexylamine; DIEA, N,N-diisopropylethylamine; DMAP, 4-dimethylaminopyridine; DMF, N,N-dimethylformamide; DMF(Npa)₂, N,N-dimethylformamide dineopentyl acetal; Dts, dithiasuccinoyl; Fmoc, 9-fluorenylmethyloxycarbonyl; HOBt, 1-hydroxybenzotriazole; Pbs, title handle of this paper; Pyr*Tos, pyridinium tosylate; (R, polystyrene resin support; TFAA, trifluoroacetic anhydride; THF, tetrahydrofuran; Tmse, 2-trimethylsilylethyl
- (a) G. Barany and R.B. Merrifield, J. Am. Chem. Soc., 99, 7363-7365 (1977).
 (b) G. Barany and F. Albericio, J. Am. Chem. Soc., 107, 4936-4942 (1985).
 (c) F. Albericio, U. Syomczynska, D.G. Mullen, S. Zalipsky, and G. Barany in U. Ragnarsson, ed., Peptides 1984, Proceedings of the Eighteenth European Peptide Symposium, Almqvist and Wiksell, Stockholm, Sweden, 1984, pp. 181-184.
- (a) R.B. Merrifield, J. Am. Chem. Soc., 85, 2149-2154 (1963).
 (b) G. Barany and R.B. Merrifield in E. Gross and J. Meienhofer, eds., <u>The Peptides</u>, Vol. 2, Academic Press, New York, 1979, pp. 1-284.
 (c) G. Barany, N.K. Cordonier, and D.G. Mullen in M. Grayson and J.I. Kroschwitz eds., <u>Encyclopedia of Polymer Science</u>, 2nd Ed., John Wiley and Sons, New York, in press, 1987.

- 6. An orthogonal system is defined as one using two or more independent classes of protecting groups that are removable by differing chemical mechanisms. Consequently, these groups can be selectively removed in any order in the presence of the other classes, see refs 4a, 5b.
- 7. A handle is defined as a bifunctional spacer that serves to link the first amino acid to the resin in two discrete steps, see refs 5b and 5c for reviews and additional examples.
- 8. (a) A.R. Mitchell, S.B. Kent, M. Engelhard, and R.B. Merrifield, <u>J. Org. Chem.</u>, 43, 2845-2852 (1978).
 (b) R.C. Sheppard and B.J. Williams, <u>Int. J. Peptide Protein Res.</u>, 20, 451-454 (1982).
 (c) M. Mutter and D. Bellof, <u>Helv. Chim. Acta</u>, 67, 2009-2016 (1984).
 (d) F. Albericio and G. Barany, <u>Int. J. Peptide Protein Res.</u>, 23, 342-349 (1984).
 (e) F. Albericio and G. Barany, <u>Int. J. Peptide Protein Res.</u>, 26, 92-97 (1985).
- 9. The idea of using fluoridolysable silicon-containing protecting groups for organic synthesis was introduced by E.J. Corey and A. Venkateswarlu, J. Am. Chem. Soc., 94, 6190-6191 (1972).
- For previous applications of fluoridolysable protecting groups to peptide synthesis:

 (a) P. Sieber, Helv. Chim. Acta, 60, 2711-2716 (1977).
 (b) L.A. Carpino, J.-H. Tsao, H. Ringsdorf, E. Fell, and G. Hettrich, J. Chem. Soc., Chem. Commun., pp. 358-359 (1978).
- 11. (a) D.S. Kemp and D.C. Roberts, <u>Tetrahedron Lett.</u>, pp. 4629-4632 (1975). (b) J.P. Tam, R.D. DiMarchi, and R.B. Merrifield, <u>Int. J. Peptide Protein Res.</u>, 16, 412-425 (1980).
- 12. S.S. Wang, J. Am. Chem. Soc., 95, 1328-1333 (1973).
- 13. L.A. Carpino and G.Y. Han, J. Org. Chem., 37, 3404-3409 (1972).
- 14. All organic compounds reported in the scheme are new and gave satisfactory elemental analysis and spectroscopic data.
- 15. J.V. Crivello, J. Org. Chem., 46, 3056-3060 (1981).
- 16. M. Miyashita, A. Yoshikoshi, and P.A. Grieco, J. Org. Chem., 42, 3772-3774 (1977).
- 17. S. Ram and R.E. Ehrenkaufer, Tetrahedron Lett., pp. 3415-3418 (1984).
- 18. Earlier trials using succinic anhydride were abandonded when it was found that the subsequent esterification step with 2,4,5-trichlorophenol was accompanied by succinimide formation.
- 19. The crude protected peptide was found to be essentially pure by HPLC: C-18 column; linear gradient starting from 0.01% aqueous HCl taken over 20 min to neat CH₃CN; flowrate 0.9 mL/min; a single peak eluted at 8.9 min. Fast atom bombardment mass spectrometry gave the expected M+H molecular ion at 459 amu.
- This protected peptide has been made previously: (a) V.K. Sarin, S.B. Kent, A.R. Mitchell, and R.B. Merrifield, J. Am. Chem. Soc., 106, 7845-7850 (1984).
 (b) F-S. Tjoeng, W. Staines, S. St.-Pierre, and R.S. Hodges, <u>Biochim. Biophys. Acta</u>, 490, 489-496 (1977).
- 21. J.M. Manning and S. Moore, J. Biol. Chem., 243, 5591-5597 (1968).

(Received in USA 29 October 1986)